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# Shoot growth and anthesis in Vitis1)

bу

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# La croissance des rameaux et l'anthèse de la vigne

Résumé. — La majorité des rameaux verts de la vigne montre un nombre assez constant d'entrenocuds au moment de l'anthèse. Vitis vinifera L. (16—19) présente un plus grand nombre d'entrenocuds que V. labruscana Baller (12—14). On retrouve dans diverses conditions (années, variétés, pays, climats, pratiques culturales) cette synchronisation de la vitesse du développement des noeuds de l'apex végétatif avec celle des fleurs sur les inflorescences. La vigueur de la vigne elle-même est le facteur responsable des plus grandes variations dans le nombre d'entrenoeuds au moment de l'anthèse.

#### Introduction

Grapevine shoot development has been divided into 4 morphological and chronological phases: (a) formation of nodes in the buds which become the dormant compound bud at the end of the first growing season; (b) formation of new nodes and elongation of all internodes from bud swell (Baggodini 1952) to anthesis in the spring of the next growing season; (c) further shoot growth, especially elongation of younger internodes, from flowering to midsummer; (d) slow growth and maturation of the shoot into a cane, from mid-summer to leaf abscission (Bugnon and Bessis 1968, Bernard 1974, 1975). Inflorescences form during phase (a) and flowers during phase (b) (Carouds 1970, 1971, Scholefield and Ward 1975); fruit set and berry development occur during phases (c) and (d).

We had noted that at phase (b) grape shoots appeared to have a constant number of internodes. The preliminary data in Fig. 1 suggest that Vitis vinifera L. cv. Muscat Gordo Blanco in South Australia flowered when the shoots had an average of 17 visible internodes despite large differences in time of flowering and shoot length. We therefore investigated the morphological relationships of shoot growth and anthesis during phase (b) in several cultivars of V. vinifera and V. labruscana Balley growing in Australia and New York, U.S.A. Some observations were also made in Switzerland.

## Materials and methods

There are at least two categories of shoots on most grapevines: the major and most obvious group is longer and has more nodes at the end of phase (d) in compar-

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ison with the shorter shoots of the smaller group (up to 30% of all shoots on a vine). Both groups may emerge from primary buds and may bear inflorescences. Our studies were confined to the larger group because they seemed more expressive of the growth potential of shoots of cultivated grapevines.

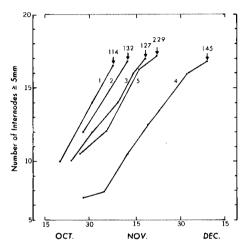


Fig. 1: Mean number of internodes per shoot of V. vinifera cv. Muscat Gordo Blanco in 3 vineyards in South Australia (1, 2 — Waikerie 1963, 1964; 3, 4 = Nuriotpa 1963, 1964; 5 = Waite 1964). Values are based on measurements of 20 to 30 shoots. The date of anthesis (↓) and the average length of the shoot (cm) on that date are indicated.
 Nombre moyen d'entrenoeuds des rameaux chez V. vinifera cv. Muscat Gordo Blanco.

Nombre moyen d'entrenceus des rameaux chez v. mujera cv. mustat controlle dans 3 vignobles en Australie du Sud (1, 2 = Waikerie 1963, 1964; 3, 4 = Nuriootpa 1963, 1964; 5 = Waite 1964). Valeurs basées sur l'observation de 20—30 rameaux verts. Sont indiquées la date de l'anthèse ( \( \psi \)) et la longueur moyenne des rameaux (cm) à cette date.

The 8 cultivars were examined at 3 different climatic sites (Tables 1 and 2). Vines in South Australia were spur-pruned and trained to a Claremont trellis (Course 1974), and those in New York were cane-pruned and trained to Hudson River Umbrella (Shauls et al. 1972). White Riesling vines in 1975 in New York showed so much winter injury (Shauls et al. 1968) that it was difficult to select shoots that continued to grow throughout the flowering period.

About 25 actively growing, well exposed shoots per cultivar were randomly selected before flowering began at each site in South Australia and New York. Shoot measurements were made at 1-2-day intervals during the course of flowering. The number and lengths of internodes were determined along each shoot from the most basal internode  $\geqslant 10$  mm long to the most apical internode  $\geqslant 5$  mm. At the same time the percentage of flowers at anthesis (i. e., with caps (calyptras) abscised) on all clusters on each shoot was estimated. These data were plotted against time and the date of 70% anthesis was determined. This stage was selected because the rate of change was then rapid and hence the interpolated date had small variability

(Table 2). This date was applied to graphs of internode number and shoot length against time to determine values at 70% anthesis.

Climatic data from weather stations near sites at which grape shoots were measured') Données climatiques des stations météorologiques près des sites exnérimentaux Table 1

		Tempera	Temperature (°C)		Ave.	Avg. relative	Avg. annu	Avg. annual precipitation
Site	Avg	Avg. daily	Abs	Absolute	humic	humidity (%)	Sum of	Avg. number
	Max.	Min.	Max.	Min.	A.M.	P. M.	- monthly avg. (mm)	of days with ≥ 0,3 mm
Waite Institute, Adelaide, South Australia	20.9	11.9	44.3	0.3	61	48	629	135
Albany, New York U.S.A.	13.9	4.4	40.0	—32.2	78	62	879	137
Zürich Switzerland	14.0	5.3	37.7	-23.2	98	99	1089	158

Apical buds covered by the lowest pair of stipules and containing internodes < 5 mm were taken from 7—10 similar shoots at some sites during the flowering period. Under a stereoscopic microscope all emerged leaf primordia were counted to determine the number of internodes then initiated. The total length of the bud, negligible in relation to the entire shoot length, was not included in the shoot length.

Table 2

Longueur et nombre d'entrenoeuds des rameaux verts des vignes quand l'anthèse est réalisée a 70% Shoot lengths and internode numbers of Vitis species at 70% anthesis

					Shoot length	Number	Number of internodes	Number of shoots
Site	Species	Cultivar	Year	Date	(cm)	Visible	In shoot tip*)	measured
Adelaide	V minifera	Grenache	1974	Nov. $27.6 \pm 0.6$	104 ± 3	$19.2 \pm 0.3$	7.5 ± 0.2	23
South			1975	Nov. $17.6 \pm 0.2$	$108 \pm 3$	$18.7\pm0.3$	I	22
Anstralia		Shiraz	1974	Nov. 25.8 $\pm$ 0.5	$132 \pm 5$	$18.1\pm0.2$	$8.8 \pm 0.2$	23
rasii ana			1975	Nov. $17.6 \pm 0.2$	$143\pm4$	$18.9\pm0.2$	I	25
		Cahomot	1974	Nov. $23.2 \pm 0.4$	90 ± 4	$16.6\pm0.2$	$7.5\pm0.2$	24
		Sauvienon	1975	Nov. $16.7 \pm 0.3$	$108 \pm 3$	$17.5\pm0.3$	!	26
		White Riesling	_	Nov. $14.8 \pm 0.2$	$72 \pm 3$	$16.4\pm0.1$	l	26
Geneva	V minifera	White Riesling 1975	1975	June 21.0 ± 0.3	67 ± 6	$15.9 \pm 0.7$	$9.5 \pm 0.3$	22
New York			1976	June 23.2 $\pm$ 0.2	80 ± 4	$18.0\pm0.2$	$10.8 \pm 0.2$	24
TOT HOLD	V. lahruscana	Concord	1975	June $15.7 \pm 0.1$	87 ± 5	$11.5\pm0.2$	$7.6 \pm 0.3$	20
			1976	June 17.8 $\pm$ 0.1	$104 \pm 4$	$13.5\pm0.2$	$8.0 \pm 0.3$	22
Wädenswil.	V. vinifera	Chasselas	1976		83 ± 4	$16.2 \pm 0.5$	1	ı
Switzerland	•	Pinot noir	1976	1	$94 \pm 3$	$14.5 \pm 0.5$	1	1
	V. labruscana	I	1976	1	ı	12.0	I	1

ny visible intermodes include those >> 10 mm long at the base of the shoot to those >> 5 mm long at the tip.

3) Intermodes <> 5 mm include all those leaf primordia in the apical bud covered by the stipules of the lowest folded leaf.

This measurement was made only once during flowering, with the assumption that these values did not significantly change during the flowering period.

Shoot measurements at Wädenswil, Switzerland, were made on 24th June 1976 using a less exact method. 60 shoots each of Pinot Noir (clone 2/10) and Chasselas (syn. Weißer Gutedel) growing on 5 C rootstock on a "Zweietagen" system (Eggenberger et al. 1975) at the Federal Grape Station were assessed for internode number, shoot length, and percentage of anthesis. These figures were grouped into 5 or 6 classes based on stage of flowering, the averages and standard errors of the means of each calculated, and the values at 70% anthesis interpolated. On one vine of V. labruscana cv. unknown 7 shoots were assessed in the same manner.

To gain information on one possible source of variation in the internode number at flowering, shoots from vines showing different rates of shoot elongation were compared in 1975. Three 10-year-old Muscat Gordo Blanco vines at Adelaide were selected for their widely different shoot vigour brought about mainly by root restriction: the weak vine grew in a glazed pipe (length 53 cm, internal diameter 31 cm); the other 2 vines grew in a vineyard, the more vigorous vine being in an outside row. On each vine internodes of 3 shoots were measured on 4 dates, twice before and twice after 70% anthesis.

To determine the number of nodes formed during period (a), 21 primary buds from the lower 5 nodes of White Riesling canes in New York were dissected at bud swell in 1975, and the number of internodes assessed.

## Results

The South Australian sites differed from the New York and Swiss sites in having hotter, drier summers and winter temperatures above freezing (Table 1). Irrigation was used in Australia to bring the annual precipitation up to about 760 mm (COOMBE 1974).

A striking feature of the shoot measurements is the low variability of the number of internodes per shoot at 70% anthesis (Table 2). The standard errors of the means are only 1.0 to 1.7% for all South Australian and New York data, except for the greater variability (4.4% of the means) of White Riesling in 1975 in New York. In contrast, the shoot lengths between sites and cultivars varied widely. The standard errors of the means of shoot lengths were 3 to 6% of the means, except for the greater variability (9%) of White Riesling in 1975 in New York.

It is clear that the mean numbers of internodes at 70% anthesis were consistently higher for V. vinifera (16—19) than for V. labruscana (12—14). The average internode number for V. labruscana was similar in Switzerland and New York. The values for V. vinifera were generally lower and more variable in New York and Switzerland than those for the same species in South Australia (Table 2).

The internode lengths of the 3 shoots from each of the Muscat vines of different vigour were similar and only one from each vine is shown in Fig. 2. The shoot lengths on 19th December indicate the large vigour differences in the 3 vines: 54, 141 and 243 cm. The number of internodes at 70% anthesis were 16.5, 18 and 22, respectively. Thus, although the variation within each vigour group was small, the differences between the groups were relatively large, in fact larger than the differences between V vinifera cultivars growing in 3 countries (Table 1). It must be pointed out, however, that these differences in vigour were extreme. The differences in vigour were evident early in the growth of the shoot: the length of the internode above the basal

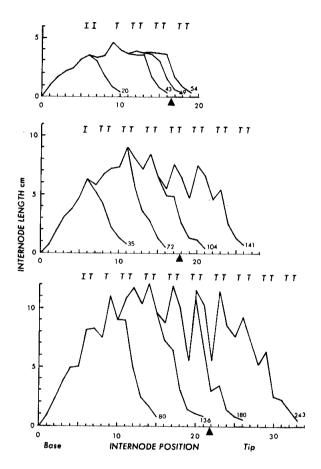


Fig. 2: Lengths of internodes on 4 dates (11th, 19th November; 1st, 19th December 1975) of a typical shoot on each of 3 vines of different vine vigour of V. vinifera cv. Muscat Gordo Blanco. The positions of inflorescences (I) and tendrils (T), the total shoot length (cm) on each date, and the internode number at 70% anthesis (\(^{\text{A}}\)) are indicated.

Top: weak vine; middle: normal vine; bottom: vigorous vine.

Longueurs des entrenoeuds d'un rameau sur 3 vignes de vigueurs différentes à 4 dates (11 et 19 novembre; ler et 19 décembre 1975) chez V. vinifera cv. Muscat Gordo Blanco. Position de l'inflorescence = I; viille = T; nombre d'entrenoeuds à 70 % anthèse = A; longueur totale du rameau à chaque date en cm. En haut: vigne faible; milieu: vigne normale; en bas: vigne vigoureuse.

inflorescence, which had attained its full size several weeks before flowering, appeared to forecast the total growth of the shoot by the time of anthesis because it was proportional to both mean internode length and node number.

The numbers of internodes within the swelling buds of White Riesling in New York, 1975, ranged from 6.2 to 8.3, depending on nodal position.

#### Discussion

The well developed shoots on a grapevine have a relatively constant number of visible internodes when anthesis occurs, even under widely different growing conditions. This number is greater in *V. vinifera* cultivars than in *V. labruscana*: we found means of 16—19 externally visible internodes at anthesis, in agreement with the values found by BOINOVA-BONEVA (1976) in Bulgaria for *V. vinifera* cultivars, and 12—14 for *V. labruscana* cultivars. With *V. vinifera*, vigour of the individual vine (Fig. 2) influenced internode number at flowering more than did cultivar, district and cultural conditions (Table 2).

The development of the shoot involves both node formation at the apical meristem and clongation of the internodes. The nodes formed up to the stage of anthesis comprise 2 categories: a) The basal nodes, about 8 in number, were present before bud break and were initiated within the primary bud during the first growing season (Bannard and Thomas 1933, May 1964). The number of nodes varies with position of the bud along a cane (Bugnon and Bessis 1968, Carolles 1970, Buttrose 1974) or cultivar (Bouard and Pouget 1971). It is similar in *V. vinifera* and *V. labruscana* (Pratt 1959). Some of these nodes bear inflorescences. The elongation of these internodes is completed before anthesis (Fig. 2), as shown by Jacquinet and Simon (1971). There is often a short internode (Carlies et al. 1964, Jacquinet and Simon 1971) or a disruption of the 1-1-0 pattern of *V. vinifera* inflorescences and tendrils between this group of nodes and those which follow, as seen in Fig. 2 (Jacquinet 1974). b) Nodes initiated between bud break and anthesis include about 4 (*V. labruscana*) or 10 (*V. vinifera*) visible nodes and about another 8 at the shoot tip.

These results suggest that there is synchrony in the rates of development of new nodes and of flowers during period (b). It is possible that this synchrony is exercised by a dependence of the development of one organ on the development of the other. This could be tested by excision experiments. Another explanation is that the development of both may be influenced directly by the same factor; it would appear that this factor would not involve the elongation of internodes except under extreme circumstances (Fig. 2). Of hormonal candidates, the most probable are cytokinins produced by the roots (Buttrose and Mullins 1968, Mullins 1968, Pool 1975, Pool and Powell 1975), and distributed by partitioning of the xylem sap between the vegetative and the floral meristems.

However, shoot growth can be greatly modified by environmental factors. Shoot length and, to a less extent, the number of visible internodes tended to be less in New York and Switzerland, where below-freezing winter temperatures occur, than in Australia, which has warmer winters. Low winter temperatures tend to injure V.vinifera more than V.labruscana by freezing of buds or canes, or both (Shauls et al. 1968). Cold injury to phloem and cambium in trunks and canes may limit or interrupt translocation during spring and affect the synchronous development of nodes and flowers. It also greatly affects shoot elongation (Pratt, unpublished data),

but this effect is minimized in the present data by the progressive selection of growing shoots of White Riesling in New York. Growing season temperatures, which are lower in New York and Switzerland than in Australia, are known to influence the rate of internode elongation (Bugnon and Bessis 1968, Jacquinet and Simon 1971). Cultural practices, especially pruning and training, differed between sites, and may also account for some variation.

#### Summary

In the majority of shoots on a grapevine the number of internodes is fairly constant at anthesis. The number of visible internodes was higher in *Vitis vinifera* L. (16—19) than in *V. labruscana* Balley (12—14). This synchrony in the rates of development of nodes at the shoot tip and of the flowers on the inflorescences was found in a wide array of conditions (time, cultivars, countries, climates, cultural practices). The factor found to cause greatest variation between vines was vine vigour.

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## Literature cited

- Anonymous, 1968 [first published 1958], 1972: Tables of temperature, relative humidity and precipitation for the world. Parts I, III. Meteorological Office, London.
- Anonymous, 1975: Biennial Rept., 1974—75, Waite Agricult. Res. Inst., Univ. Adelaide, South
- BAGGIOLINI, M., 1952: Les stades repères dans le développement annuel de la vigne et leur utilisation pratique. Rev. Romande Agricult. Viticult. Arboricult. 8, 4-8.
- BARNARD, C. and THOMAS, J. E., 1933: Fruit bud studies. II. The Sultana: differentiation and development of the fruit buds. J. Council. Sci. Ind. Res. 6, 285--294.
- Bernard, A. C., 1974: Particularités de la croissance en année chaude et sèche chez Vitis vinifera cv. Carlgnan, France Viticole 6, 3—15.
- -, 1975: Sur la croissance de la vigne. France Viticole 7, 9-14, 39-48.
- BODNOVA-BONEVA, I., 1976: Recherches sur la morphologie de la vigne. III. Dynamique [de] l'accroissement des différentes structures morphologiques du rameau vert de la vigne, en connexion avec le debut et l'évolution de[s] étapes du cycle complet organogenique (buig.). Hort. Viticuit. Sci. 13, 85-95.
- BOUARD, J. et POUGET, R., 1971: Physiologie de la croissance et du développement. In: Ribérbau-Gayon, J. et Peynaud, E. (Eds.): Traité d'Ampélographie. I, pp. 329—413. Dunod, Paris.
- Bugnon, F. et Bessis, R., 1968: Biologie de la vigne. Acquisitions récentes et problèmes actuels.

  Masson, Paris,
- BUTTROSE, M. S., 1974: Fruitfulness in grapevines: Effects of water stress. Vitis 12, 299-305.
- and Mullins, M. G., 1968: Proportional reduction in shoot growth of grapevines with root systems maintained at constant relative volumes by repeated pruning. Austral. J. Biol. Sci. 21, 1085—1101.
- CARLIS, J., ASSAI, R., MAGNY, J. et RIVAIS, P., 1964: Différences physiologiques dans les rameaux entre la partie néoformée et la partie préformée dans le bourgeon. C. R. Acad. Sci. (Paris) 259, 3348-3351.
- Cancius, M., 1970: Recherches sur l'organogenèse et l'évolution morphologique du bourgeon latent de la vigne (Vitis vinifera L. var. Merlot). Thesis, Univ. Bordeaux,
- --- , 1971: Description des stades du développement des primordia inflorescentiels durant

- l'organogenèse des bourgeons latents de la vigne (Vitis vinifera L. var. Merlot). Connaiss. Vigne Vin 2, 163—173.
- COOMBR, B. G., 1974: Performance of wine grapes on table grape trellis. Austral. Grapegrower and Winemaker 11 (124), 75—78.
- Ecgenberger, W., Koblet, W., Mischler, M., Schwarzenbach, H. und Smon, J.-L., 1975: Weinbau. Huber, Frauenfeld.
- JACQUEINBT, A., 1974: Une méthode de contrôle de la vigueur et de la croissance de la vigne. Vitis 12, 291-298.
- et Simon, J.-L., 1971: Contribution à l'étude de la croissance des rameaux de vigne. Rev. Suisse Viticult. Arboricult. 3, 131—135.
- May, P., 1964: Über die Knospen- und Infloreszenzentwicklung der Rebe. Wein-Wiss. 19, 487-485.

  Mullins, M. G., 1967: Morphogenetic effects of roots and of some synthetic cytokinins in Vitis

  tinifera L. J. Exp. Bot. 18, 206-214.
- Pool, R. M., 1975: Effect of cytokinin on in vitro development of 'Concord' flowers. Amer. J. Enol. Viticult. 26, 43-46.
- --- and Powell, L. E., 1975: The influence of cytokinins on in vitro shoot development of 'Concord' grape, J. Amer. Soc. Hort. Sci. 100, 200-202.
- PRATT, C., 1959: Radiation damage in shoot apices of Concord grape. Amer. J. Bot. 46, 103—109 Scholeffeld, P. B. and Ward, R. C., 1975: Scanning electron microscopy of the developmental stages of the Sultana inflorescence. Vitis 14, 14—19.
- Shauur, N., Einset, J. and Pack, A. B., 1968: Growing cold-tender grape varieties in New York. N.Y. State Agricult. Exp. Sta., Geneva, Bull. 821.
- JURDAN, T. D. and TOMKINS, J. P., 1972: Cultural practices for New York vineyards., N.Y. State Coll. Agricult. Life Sci. Ext. Bull. 805.

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